

Nikon Training Notebook

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University of California, Riverside

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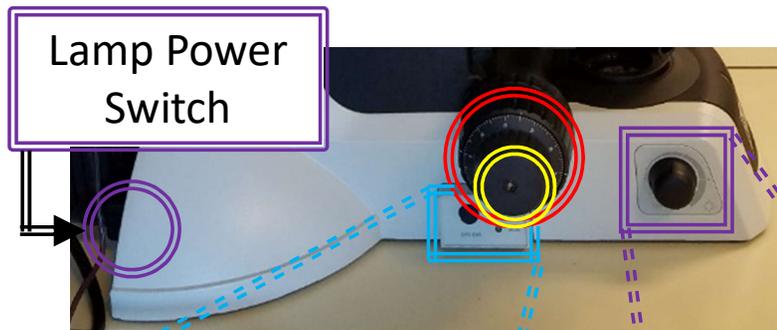
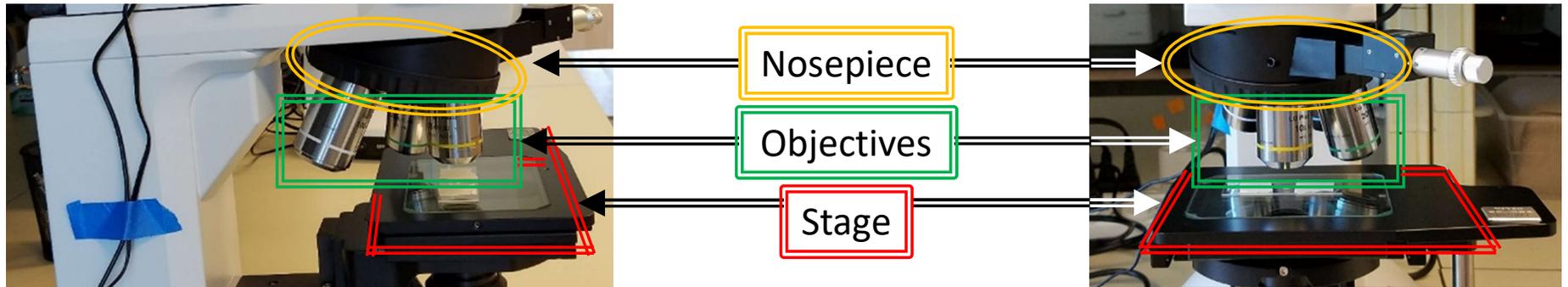
Before you begin...

- Complete the required safety training modules on UC Learning
 - Laboratory Safety Orientation (Fundamentals) 2013
 - Hazardous Waste Management
 - Compressed Gas Safety
- Submit a copy of your Training Transcript to Lab Manager
- Review the MSE Policies and Regulations
- Fill out the MSE 150, 250, 309 FAU Authorization Form with PI signature
- Provide your ENGR username to Lab Manager to set up Faces account
- Arrange a time for training with Lab Manager
- Schedule your reservation on Faces for your training

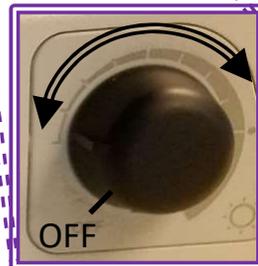
Nikon Microscope Operation

- I. Microscope Layout
- II. Startup
- III. EPI: Bright Field
- IV. EPI: Dark Field
- V. EPI: Polarization
- VI. EPI: Differential Interference Contrast (DIC)
- VII. DIA: Bright Field
- VIII. Image Capture
- IX. Cleanup
- X. ImageJ

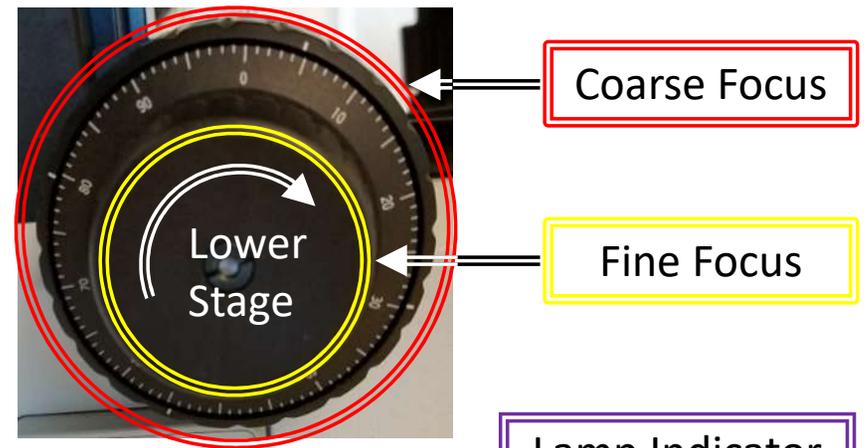
I. Microscope Layout – 2/4



EPI/DIA Selector



Lamp Brightness Knob



Coarse Focus

Fine Focus

Lamp Indicator

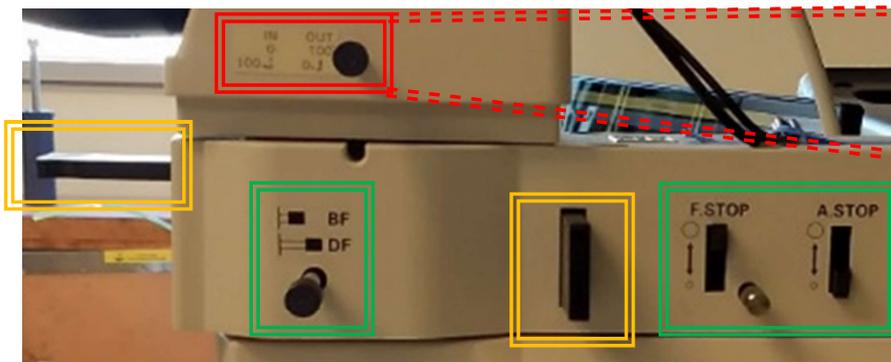
Lamp ON \Rightarrow



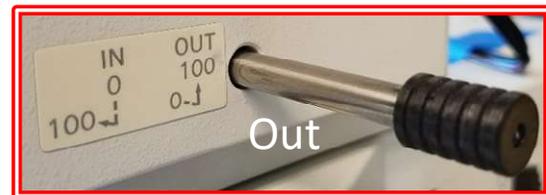
Lamp OFF \Rightarrow



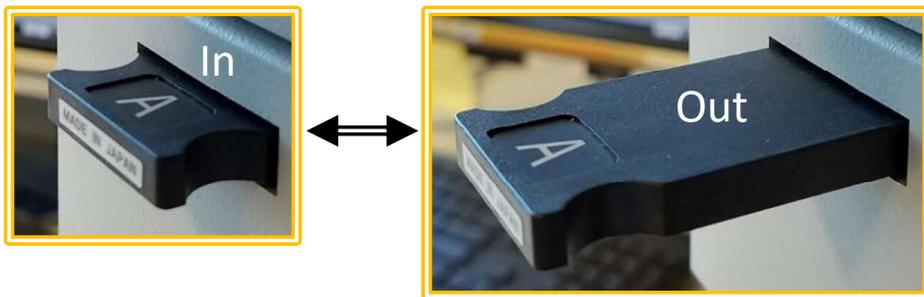
I. Microscope Layout – 3/4



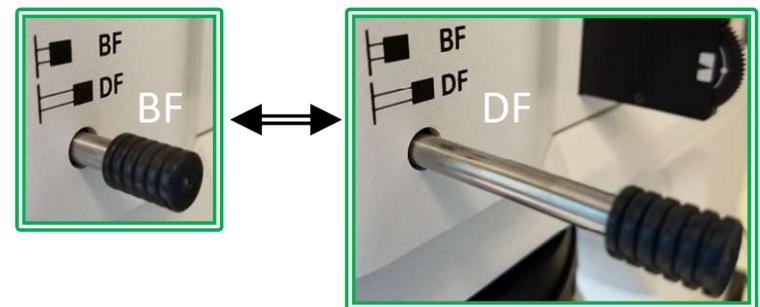
Optical Path Selector Lever



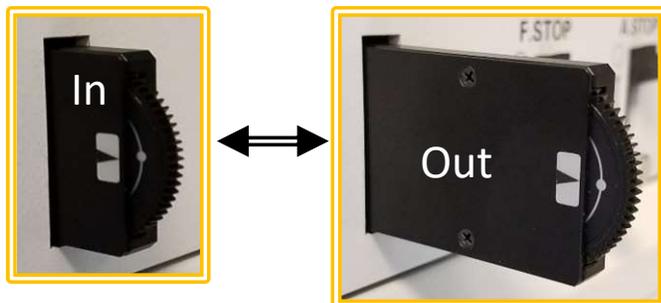
Analyzer Plate



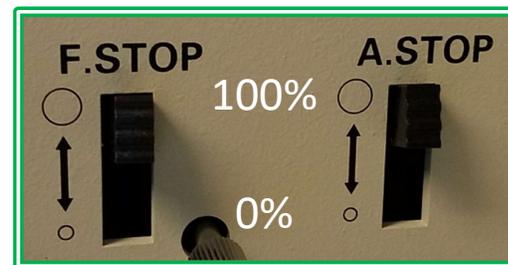
Bright field/Dark field Selector Lever



Polarizer Slider

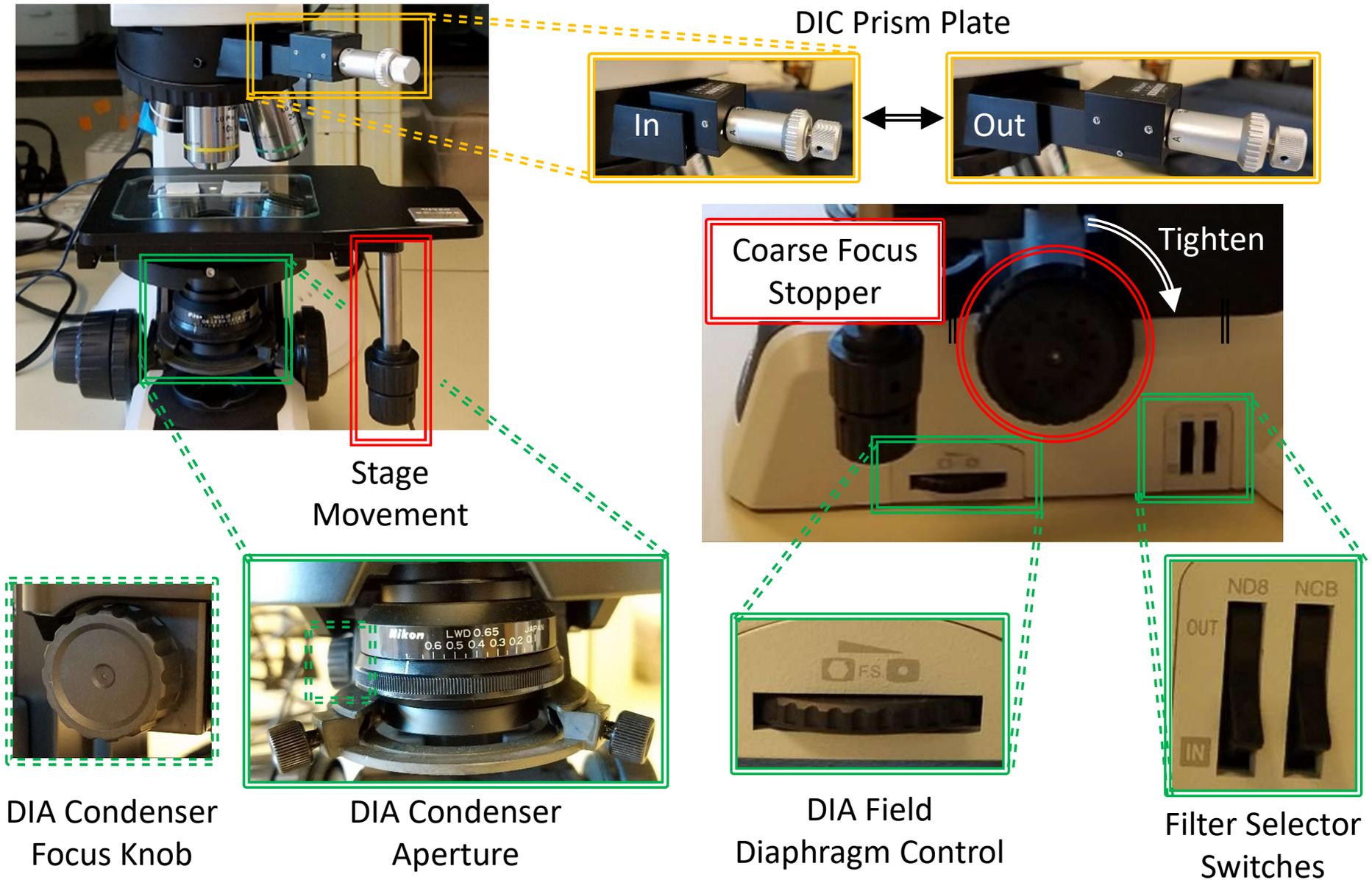


EPI Field Diaphragm Stop



EPI Aperture Diaphragm Stop

I. Microscope Layout – 4/4

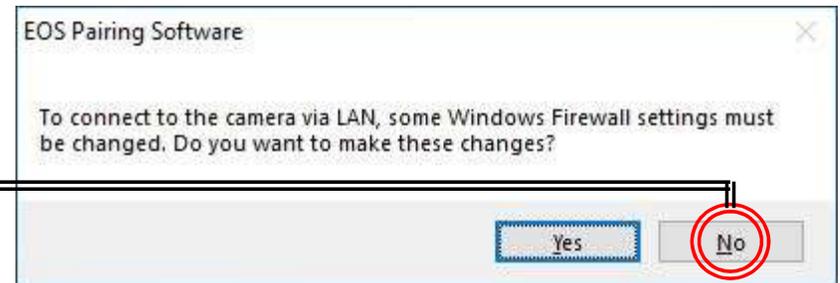


II. Startup – 1/5

1. Double-click on *EOS Utility*



2. Click **NO** when asked to change Firewall settings



3. The EOS Utility Launcher may show that the camera is not connected to the computer



4. Toggle Camera **ON/OFF** switch to connect it to the computer – keep in **ON** position

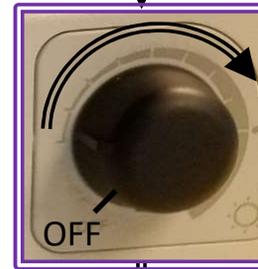


5. Click on *Camera settings/Remote shooting*



II. Startup – 3/5

9. Rotate the **Lamp Brightness Knob** until the light indicator goes from **orange (OFF)** to **green (ON)**



10. If light is missing, turn on **Lamp Power Switch** on back



11. For **Camera View**: Pull lever completely out

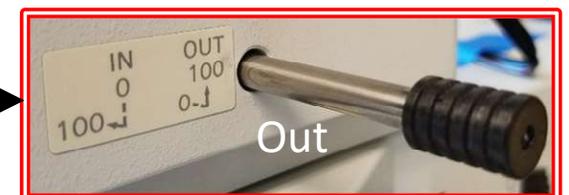
For **Binocular Eyepiece**: Push lever completely in



Optical Path Selector Lever

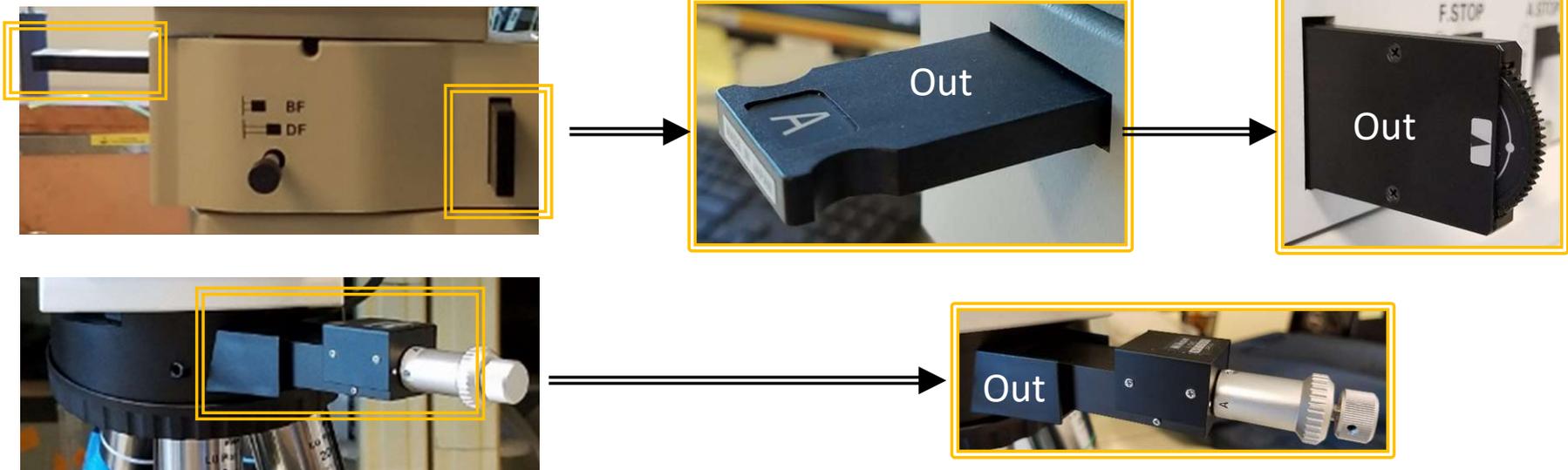
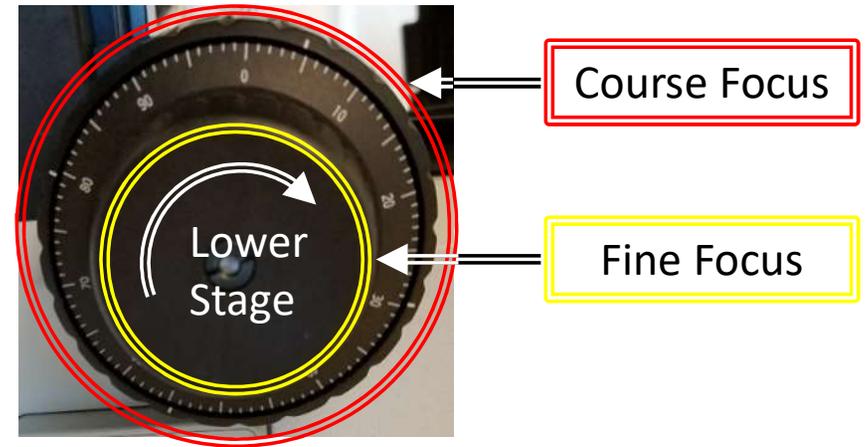
Binocular Eyepiece

Camera View



II. Startup – 4/5

12. Lower stage first by turning **Coarse Focus** knob **TOWARD** you
13. Place sample on microscope stage
14. Rotate **Nosepiece** and start with the **10X magnification** first
15. Pull out **Analyzer**, **Polarizer** and **DIC Prism** if inserted

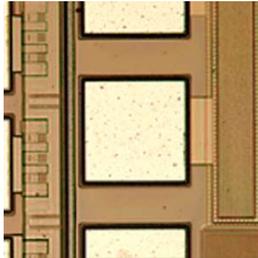


II. Startup – 5/5

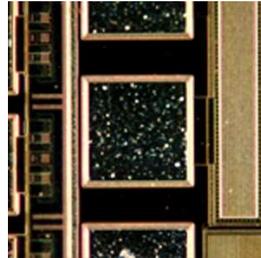
16. Identify which microscope mode you wish to use:

Episcopic Illumination ()

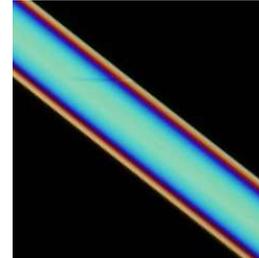
III. Bright field



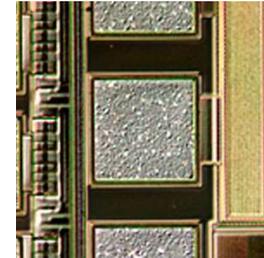
IV. Dark field



V. Polarization

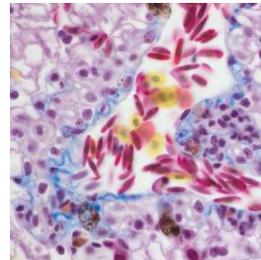


VI. Differential Interference Contrast (DIC)



Diascopic Illumination()

VII. Bright field

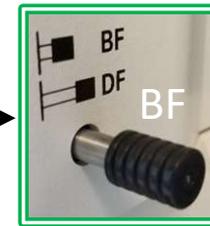
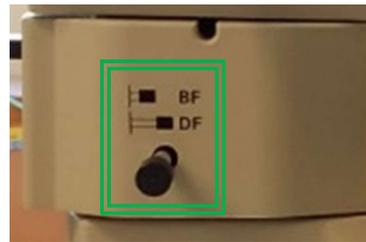


III. EPI: Bright Field – 1/3

1. Press the **EPI/DIA** selector and set to **EPI**



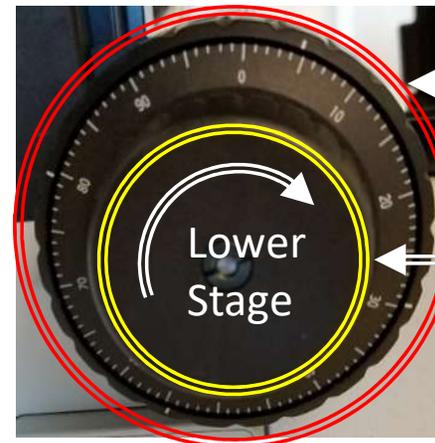
2. Push **Bright/Dark Field** selector lever to fully in **BF** position



3. Adjust the brightness with the **Brightness Control** as necessary



4. Focus on specimen by adjusting the **Coarse/Fine Focus** knobs

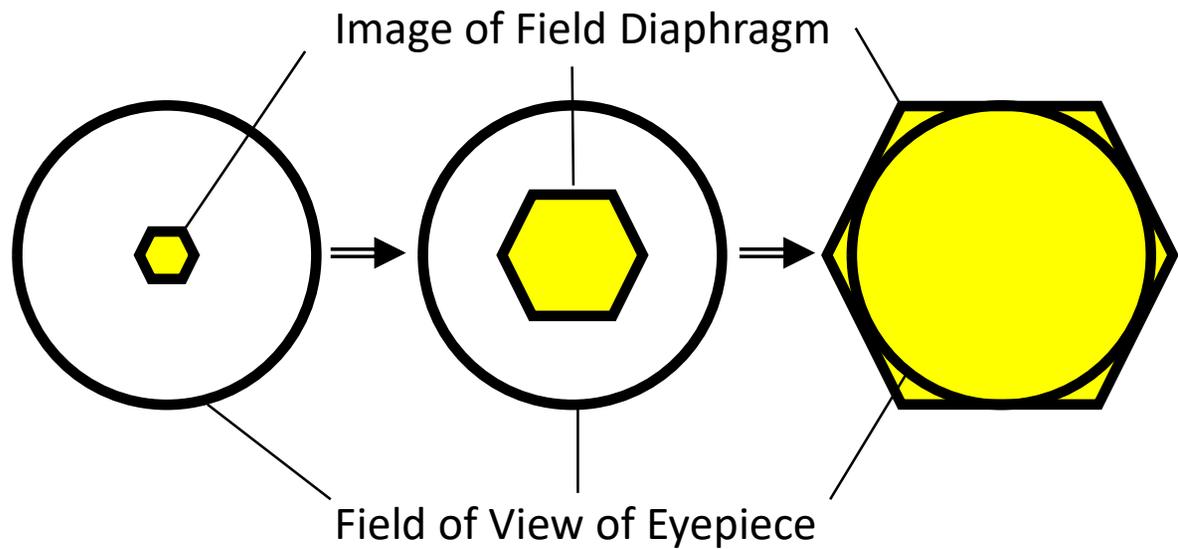
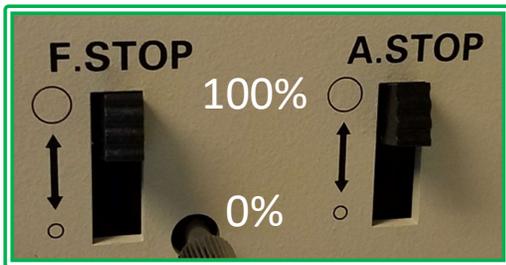


Course Focus

Fine Focus

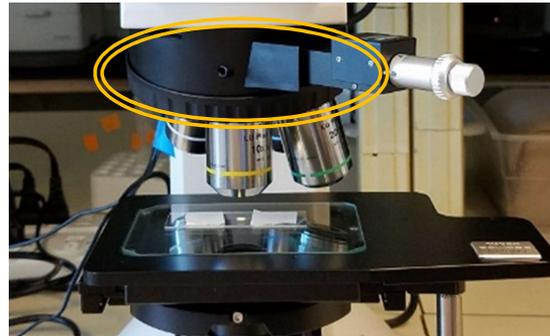
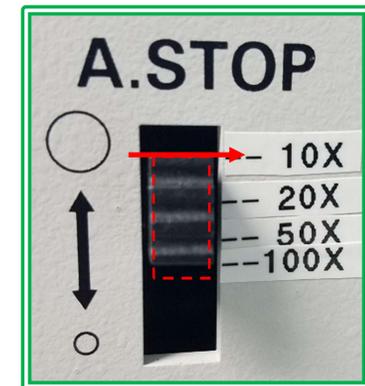
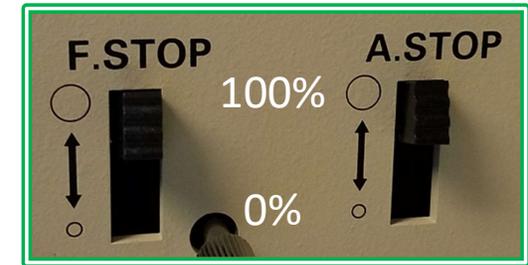
III. EPI: Bright Field – 2/3

5. Adjust the **F. STOP** (field diaphragm) by sliding levers up and down until **Image of Field Diaphragm** circumscribes the **Field of View**



III. EPI: Bright Field – 3/3

- Adjust the **A. STOP** (aperture diaphragm) by sliding levers up and down to adjust depth of field
- For each objective, recommended **A. STOP** position (**top of lever**) is shown on markings
- Switch to higher magnification objectives if desired by rotating nosepiece
- Repeat steps 3-9 until desired magnification and image quality is obtained
- Go to **Step VIII. Image Capture** when ready to acquire image

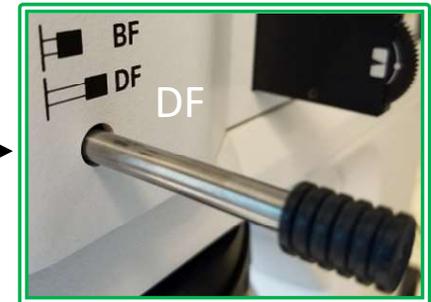
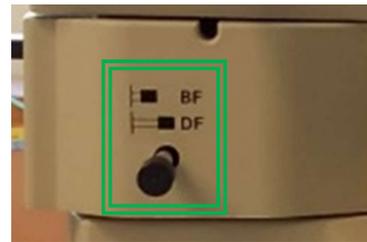


IV. EPI: Dark Field – 1/2

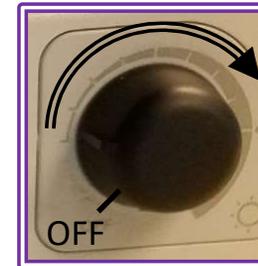
1. Press the **EPI/DIA** selector and set to **EPI**



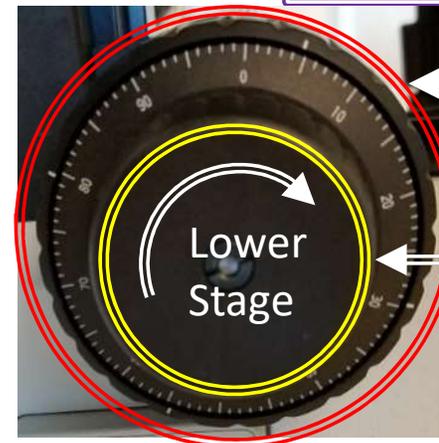
2. Pull **Bright/Dark Field** selector lever to fully out **DF** position



3. Adjust the brightness with the **Brightness Control** as necessary



4. Focus on specimen by adjusting the **Coarse/Fine Focus** knobs



Course Focus

Fine Focus

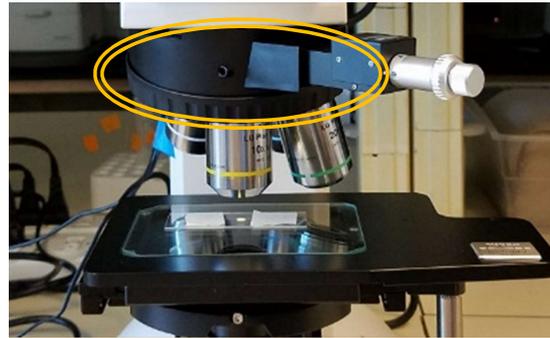
IV. EPI: Dark Field – 2/2

5. The **F. STOP** (field diaphragm) and **A. STOP** (aperture diaphragm) are automatically 100% open

Levers will have **NO** affect



6. Switch to higher magnification objectives if desired by rotating nosepiece



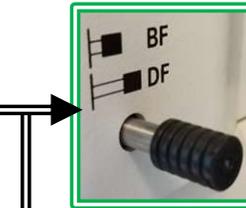
7. Repeat steps 3-6 until desired magnification and image quality is obtained
8. Go to **Step VIII. Image Capture** when ready to acquire image

V. EPI: Polarization – 1/2

1. Press the **EPI/DIA** selector and set to **EPI**



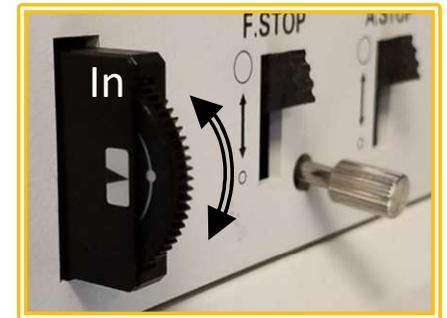
2. Adjust **Bright/Dark Field** selector lever to desired



3. Push the **Analyzer Plate** in



4. Push the **Polarizer Slider** in

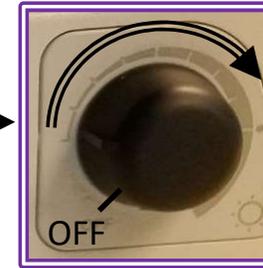


5. Rotate the polarizer to adjust the polarization from

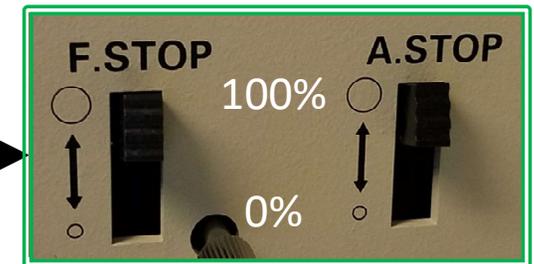
lateral  to vertical 

V. EPI: Polarization – 2/2

6. Adjust the brightness with the **Brightness Control**



7. Adjust the **F. STOP** (field diaphragm) and **A. STOP** (aperture diaphragm) by sliding levers up and down from 100% open to 0% open



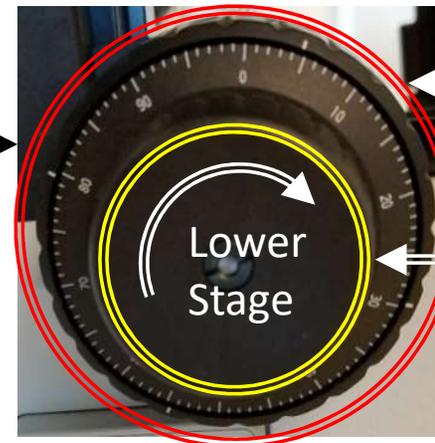
Note: **F. STOP** and **A. STOP** levers will not work if in **DF** mode

8. Focus on specimen by adjusting the **Coarse/Fine Focus** knobs

9. Switch to higher magnification objectives if desired by rotating nosepiece

10. Repeat steps 5-9 until desired magnification and image quality is obtained

11. Go to **Step VIII. Image Capture** when ready to acquire image



Course Focus

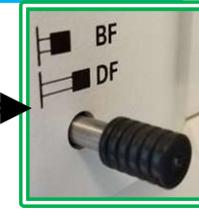
Fine Focus

VI. EPI: Differential Interference Contrast – 1/2

1. Press the **EPI/DIA** selector and set to **EPI** 



2. Adjust **Bright/Dark Field** selector lever to desired



3. Push the **Analyzer Plate** in \Rightarrow



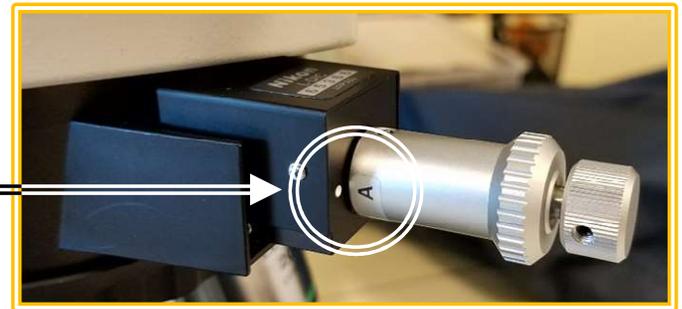
4. Push the **Polarizer Slider** in \Rightarrow



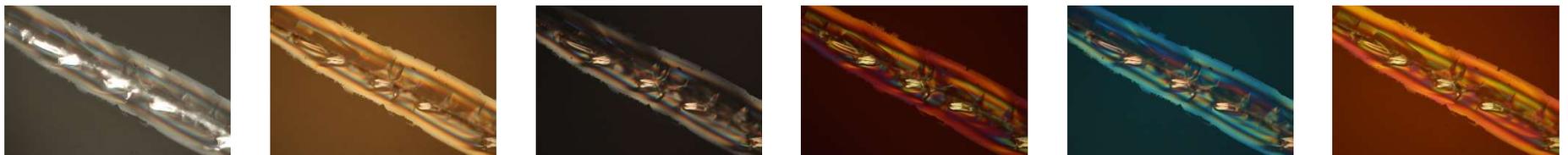
5. Rotate the polarizer to adjust the polarization from

lateral  to vertical 

6. Push the **DIC Prism** in and set to **Position A** \Rightarrow

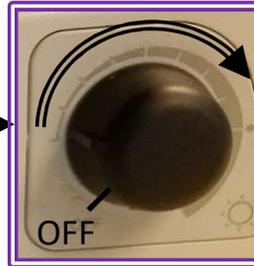


7. Rotate small knob to adjust contrast and color

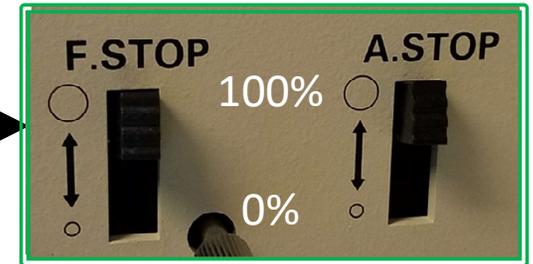


VI. EPI: Differential Interference Contrast – 2/2

8. Adjust the brightness with the **Brightness Control** ⇒



9. Adjust the **F. STOP** (field diaphragm) and **A. STOP** (aperture diaphragm) by sliding levers up and down from 100% open to 0% open ⇒



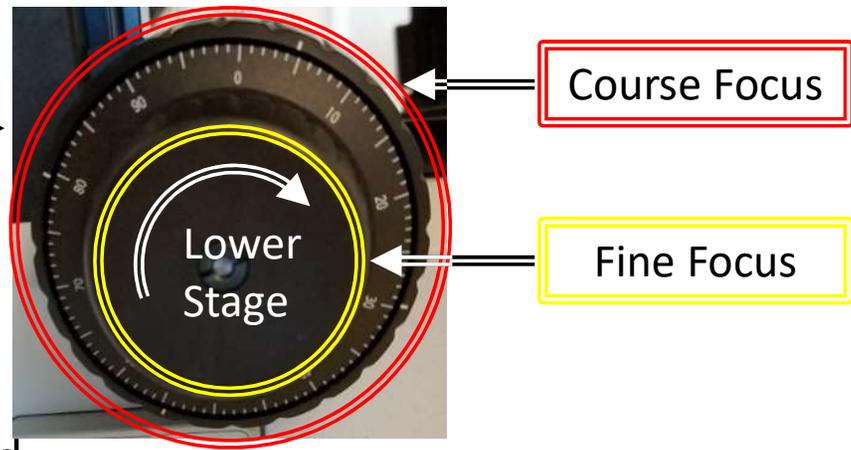
Note: **F. STOP** and **A. STOP** levers will not work if in **DF** mode

10. Focus on specimen by adjusting the **Coarse/Fine Focus** knobs ⇒

11. Switch to higher magnification objectives if desired by rotating nosepiece

12. Repeat steps 5-11 until desired magnification and image quality is obtained

13. Go to **Step VIII. Image Capture** when ready to acquire image

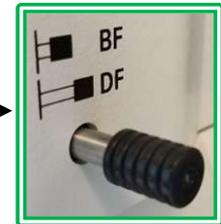


VII. DIA: Bright Field – 1/2

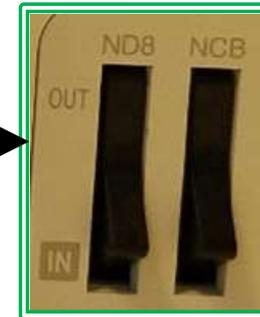
1. Press the **EPI/DIA** selector and set to **DIA** 



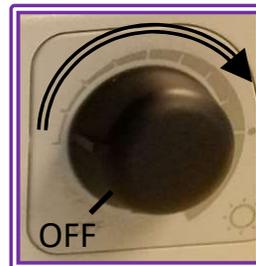
2. Push **Bright/Dark Field** selector lever to fully in **BF** position



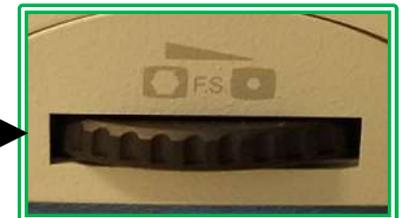
3. Select **NCB filter** (balances color) if desired



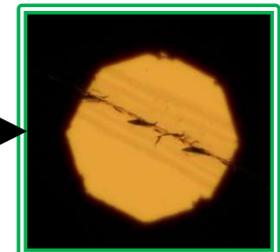
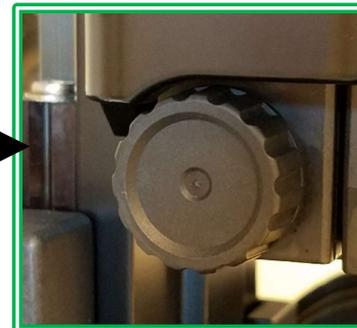
4. Adjust the brightness with the **Brightness Control**



5. Adjust the **Field Diaphragm Control** to fully closed



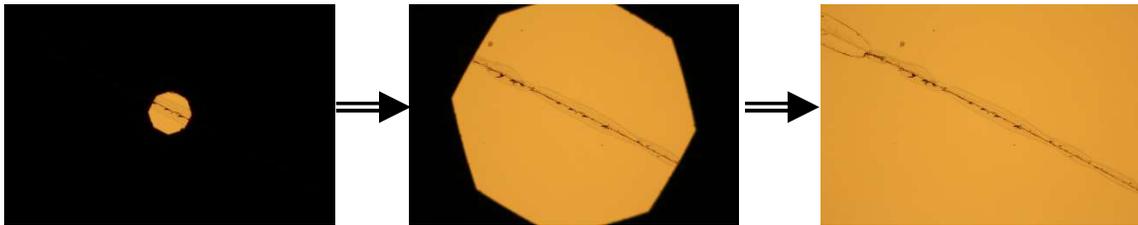
6. Adjust the **Condenser Height** until the field diaphragm is focused



VII. DIA: Bright Field – 2/2

7. Center the field diaphragm by adjusting **Centering Screws**

8. Open the **Field Diaphragm Control** until field diaphragm circumscribes the field of view



9. Focus on specimen by adjusting the **Coarse/Fine Focus** knobs

10. Adjust the **Condenser Aperture** to match **Numerical Aperture** for each objective:

10X = 0.3

20X = 0.45

50X = 0.8

100X = 0.9

11. Switch to higher magnification objectives if desired by rotating nosepiece

12. Repeat steps 3-11 until desired magnification and image quality is obtained

13. Go to **Step VIII. Image Capture** when ready to acquire image



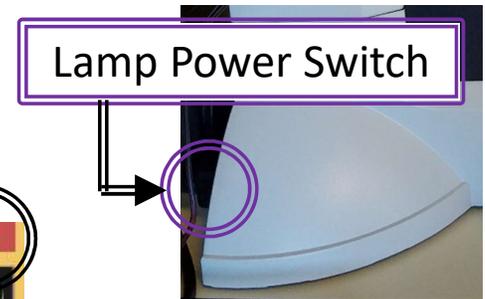
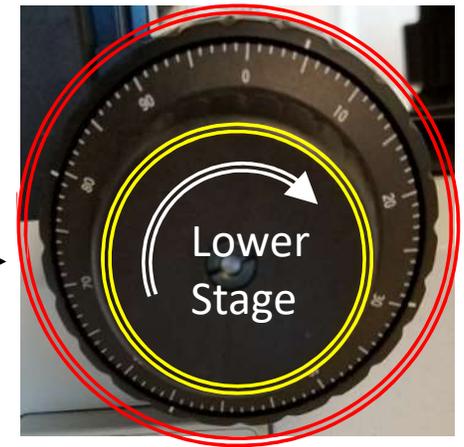
VIII. Image Capture – 1/1

1. Click on the **Folder** icon and select _____ desired folder to store saved pictures in
2. Recommend creating you own personal folder with sub-folders for each sample to help distinguish among them later
3. It is important to record the objective used for ***EACH*** image taken (necessary for scale)
4. Review **Camera Settings** _____
5. Click on the **Shutter Button** to acquire image _____



IX. Cleanup – 1/1

1. Lower the stage away from the objectives by rotating the Coarse Focus knob **TOWARD** you
2. Rotate nosepiece and place the **10x Objective** into position
3. Turn off Lamp Power Switch at the back of the microscope
4. Turn **OFF** the control software
5. **Log Out** of your ENGR account
6. Clean up and dispose of any consumables used and return any tools back to its respective containers or bins
7. Confirm that the microscope is turned **OFF** again (**NO LIGHT!**), then place cover over microscope



X. ImageJ – 1/3



1. Double-click on *ImageJ* icon

2. Click **File > Open**

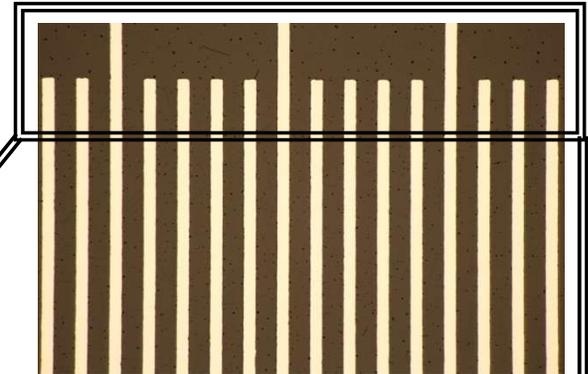


3. Locate the **Scale Bar Images** folder

4. Select the **Magnification** of the image you wish to measure (e.g. 100X) and **Open**

5. Click the **Segment Tool** and select **Straight Line**

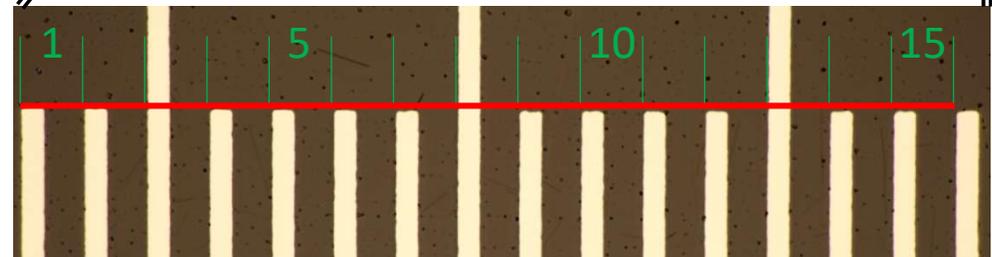
6. Draw a line that contains the maximum number of tick marks



Note: It matters where you start and end the line!

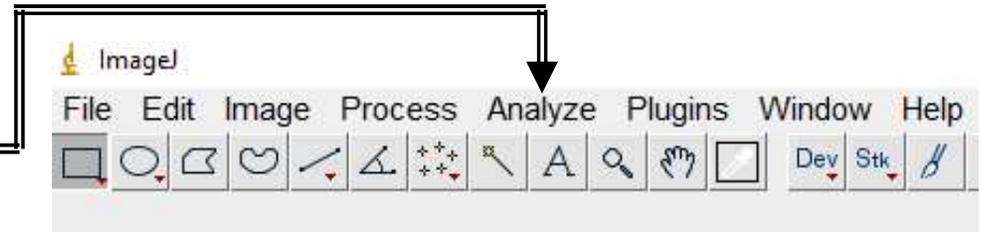
7. Count the number of tick marks contained (e.g. 15)

8. Each division is 0.01 mm (or 10 μm)



X. ImageJ – 2/3

9. Click **Analyze > Set Scale**



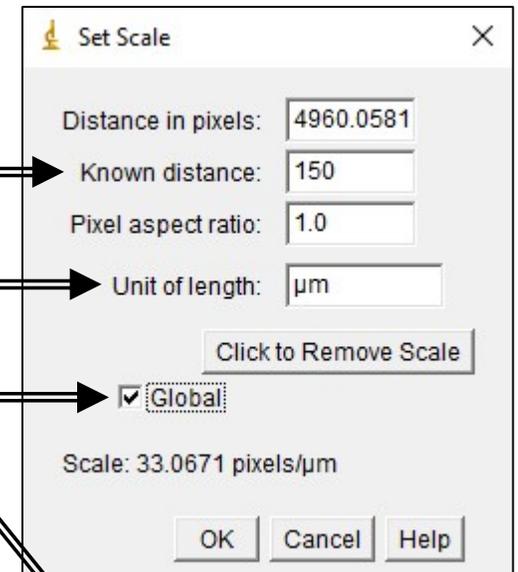
10. Enter the **Known Distance** (e.g. 150 μm) based on the number of tick marks and each division = 0.01 mm (or 10 μm)

11. Enter the **Unit of Length** to desired unit (e.g. mm)

12. Check **Global** to set scale for all images

13. Confirm your scale by drawing a new **Straight Line**

14. Click **Analyze > Measure** and check value

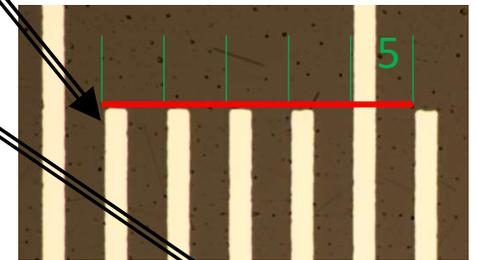


If incorrect, repeat steps 5 – 13

15. Click **File > Open** and select your image(s) of interest

16. Draw **Straight Lines** and click **Analyze > Measure**

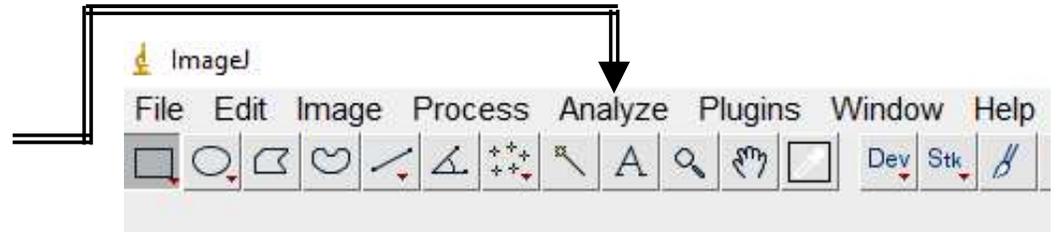
17. Repeat steps 4 – 16 for other **Magnifications**



File	Area	Mean	Min	Max	Angle	Length
1	1.508	99.107	26.987	233.667	-0.138	49.959

X. ImageJ – 3/3

9. Click **Analyze > Tools > Scale Bar**



10. Enter Width in um (e.g. 50 μm) based on the length of scale bar desired

11. Enter Height in pixels for desired scale bar thickness

12. Enter Font size for desired text size

13. Identify Color of the scale bar

14. Identify Background color (if desired)

15. Identify Location where **Scale Bar** to be placed

