Nikon Training Notebook

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Before you begin...

Complete the required safety training modules on UC Learning Laboratory Safety Orientation (Fundamentals) 2013 **Hazardous Waste Management** Compressed Gas Safety Submit a copy of your Training Transcript to Lab Manager Review the MSE Policies and Regulations Fill out the MSE 150, 250, 309 FAU Authorization Form with PI signature Provide your ENGR username to Lab Manger to set up Faces account Arrange a time for training with Lab Manager Schedule your reservation on Faces for your training

Nikon Microscope Operation

I. Microscope Layout

II. Startup

III. EPI: Bright Field

IV. EPI: Dark Field

V. EPI: Polarization

VI. EPI: Differential Interference Contrast (DIC)

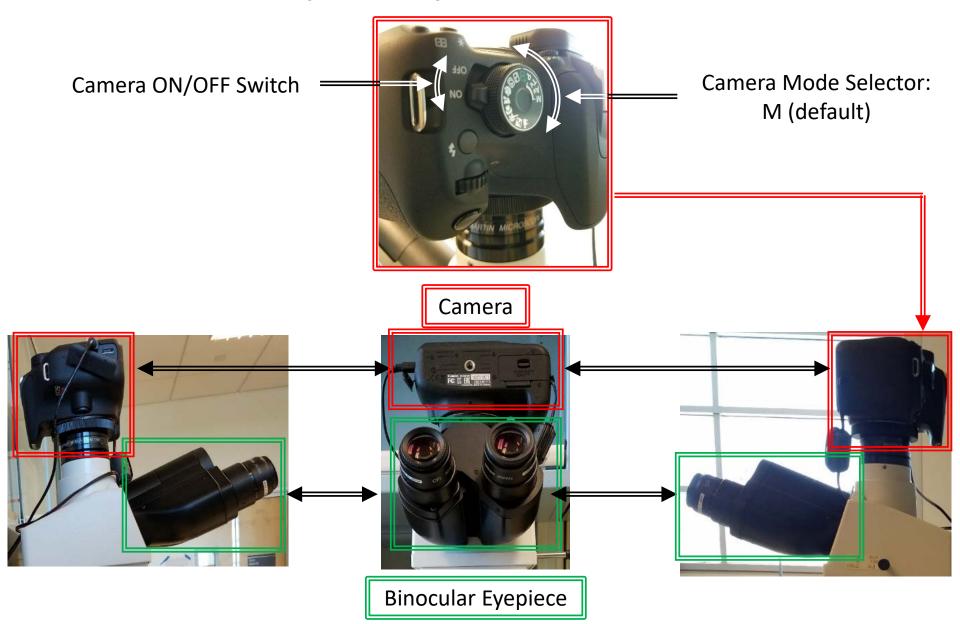
VII. DIA: Bright Field

VIII. Image Capture

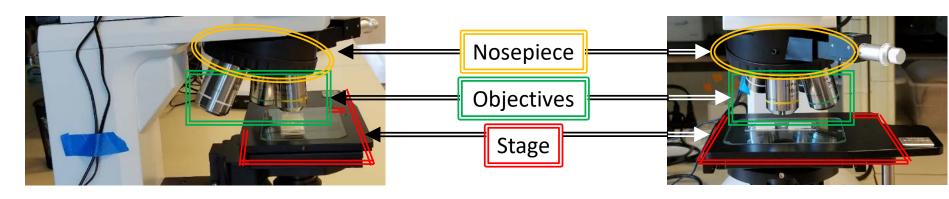
IX. Cleanup

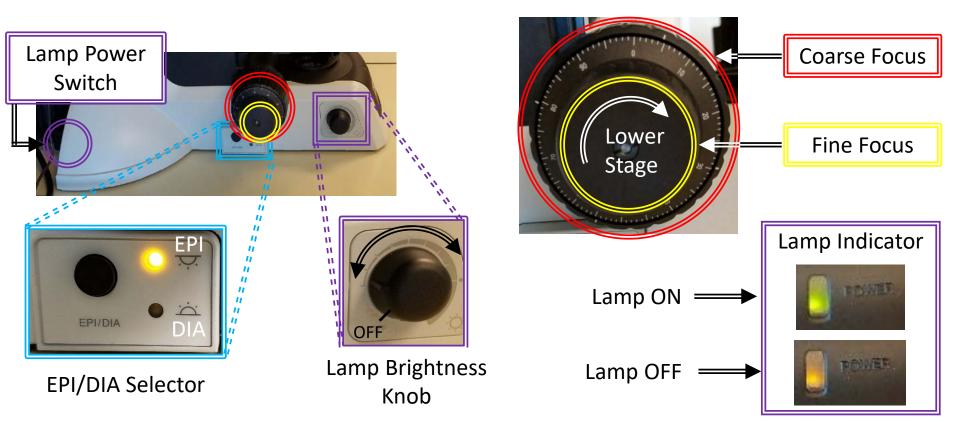
X. ImageJ

I. Microscope Layout – 1/4

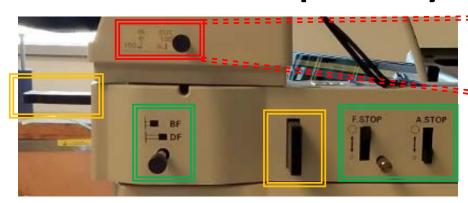


I. Microscope Layout – 2/4





I. Microscope Layout – 3/4

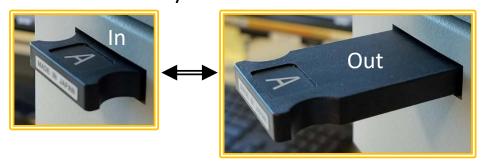




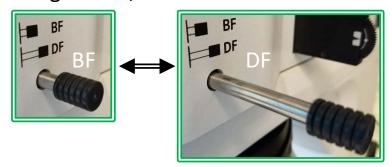
Optical Path Selector Lever



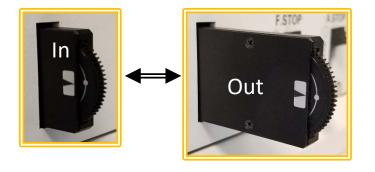
Analyzer Plate



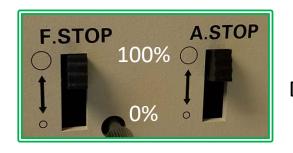
Bright field/Dark field Selector Lever



Polarizer Slider

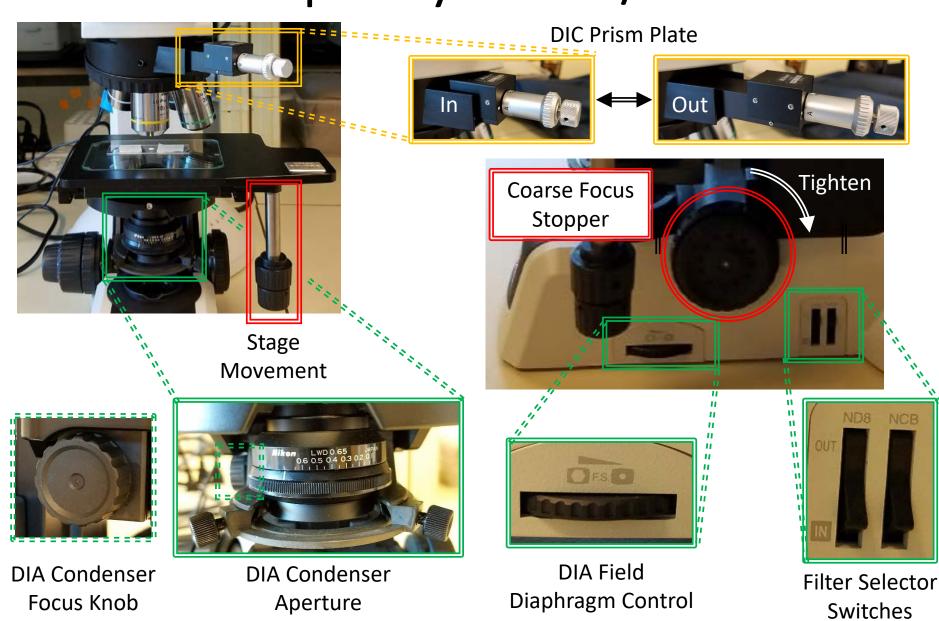


EPI Field Diaphragm Stop



EPI Aperture Diaphragm Stop

I. Microscope Layout – 4/4



II. Startup – 1/5

- 1. Sign-in to the computer with your *ENGR username* and *PW*
- 2. Double-click on *EOS Utility* icon



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Connect an EOS camera

Pairing over Wi-Fi/LAN

- 3. The EOS Utility Launcher may show that the camera is not connected to the computer
- Toggle Camera ON/OFF switch to connect it to the computer – keep in ON position



Control Camera

Starts to download images

Lets you select and download images

Camera settings/Remote shooting

Register Background Music

Preferences...

Quit

EOS Utility Launcher

5. Click on Camera settings/Remote shooting

II. Startup – 2/5

6. Confirm the following *Camera Settings* are set:

<u>Camera</u>

M = Manual



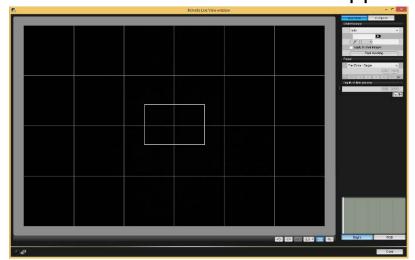
Software (right click to change)

1/50 = Shutter Speed

100 = ISO

Tungsten = Brightness

- 7. Click on *Live View shoot*
- 8. Remote Live View window will appear

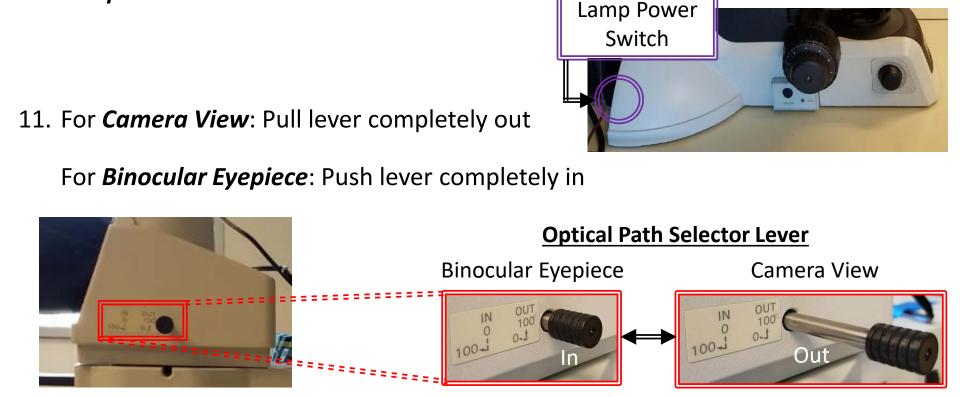




II. Startup – 3/5

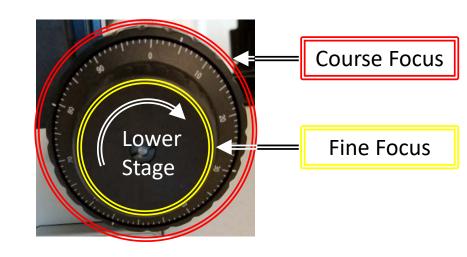
9. Rotate the *Lamp Brightness Knob* until the light indicator goes from *orange (OFF)* to *green (ON)*

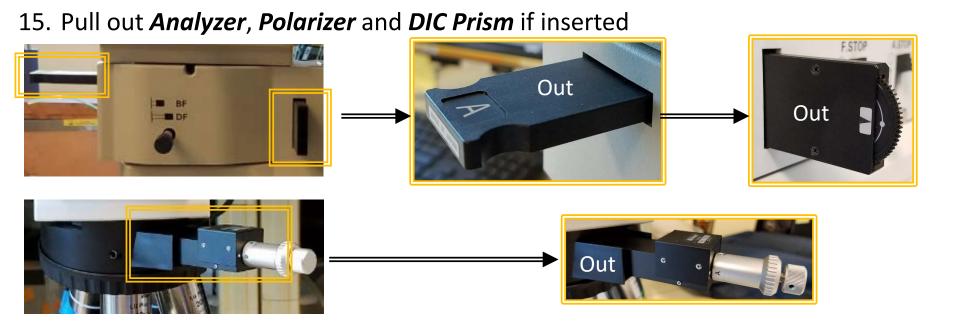
10. If light is missing, turn on **— Lamp Power Switch** on back



II. Startup – 4/5

- 12. Lower stage first by turning *Coarse Focus* knob **TOWARD** you
- 13. Place sample on microscope stage
- 14. Rotate *Nosepiece* and start with the *10X magnification* first





II. Startup – 5/5

10. Identify which microscope mode you wish to use:

Episcopic Illumination ()

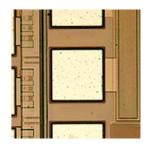


III. Bright field

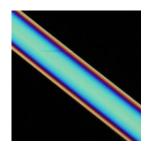
IV. Dark field

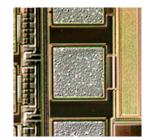
V. Polarization

VI. Differential Interference Contrast (DIC)





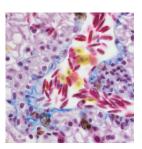




Diascopic Illumination()



VII. Bright field

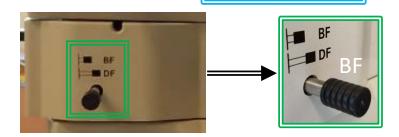


III. EPI: Bright Field – 1/3

1. Press the **EPI/DIA** selector and set to **EPI**



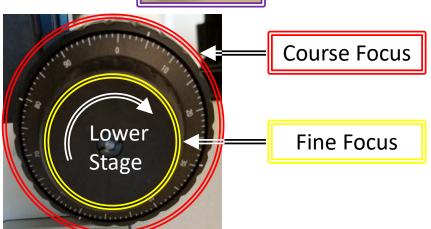
2. Push *Bright/Dark Field* selector lever to fully in *BF* position



3. Adjust the brightness with the *Brightness Control* as necessary

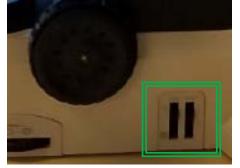


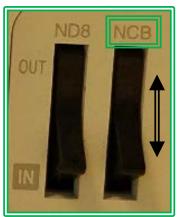
4. Focus on specimen by adjusting the *Coarse/Fine Focus* knobs



III. EPI: Bright Field – 2/3

Select if *NCB filter* (balances color) is desired:

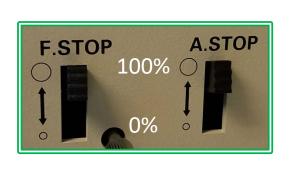


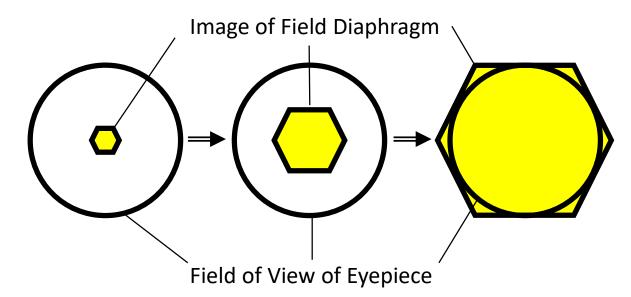






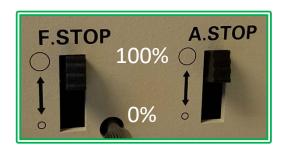
Adjust the *F. STOP* (field diaphragm)
 by sliding levers up and down until
 Image of Field Diaphragm circumscribes the *Field of View*



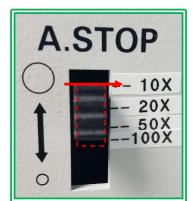


III. EPI: Bright Field – 3/3

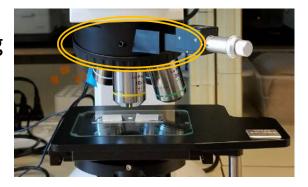
7. Adjust the **A. STOP** (aperture diaphragm) by sliding levers up and down to adjust depth of field



8. For each objective, recommended **A. STOP** position (top of lever) is shown on markings



Switch to higher magnification objectives if desired by rotating nosepiece



- 10. Repeat steps 3-9 until desired magnification and image quality is obtained
- 11. Go to *Step VIII. Image Capture* when ready to acquire image

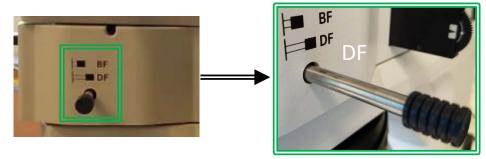
IV. EPI: Dark Field – 1/2

1. Press the EPI/DIA selector and set to EPI



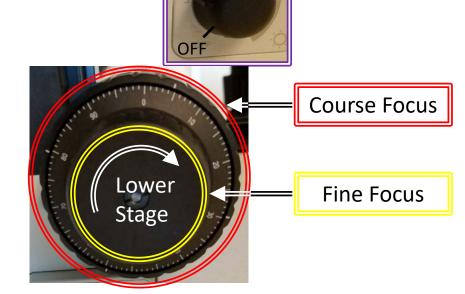


2. Pull *Bright/Dark Field* selector lever to fully out *DF* position



3. Adjust the brightness with the *Brightness Control* as necessary

4. Focus on specimen by adjusting the *Coarse/Fine Focus* knobs

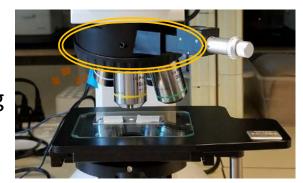


IV. EPI: Dark Field – 2/2

5. The *F. STOP* (field diaphragm) and *A. STOP* (aperture diaphragm) are automatically 100% open

Levers will have NO affect

6. Switch to higher magnification objectives if desired by rotating nosepiece



- 7. Repeat steps 3-6 until desired magnification and image quality is obtained
- 8. Go to *Step VIII. Image Capture* when ready to acquire image

V. EPI: Polarization – 1/2

Press the **EPI/DIA** selector and set to **EPI**





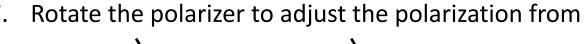
Adjust Bright/Dark Field selector lever to desired =





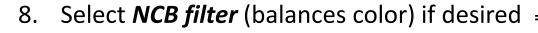
5. Push the *Analyzer Plate* in

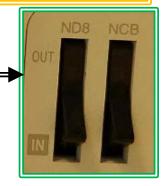












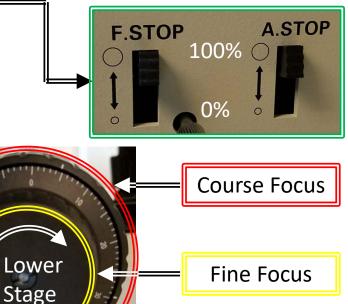
V. EPI: Polarization – 2/2

- 9. Adjust the brightness with the Brightness Control
- 10. Adjust the *F. STOP* (field diaphragm) and *A. STOP* (aperture diaphragm) by sliding levers up and down from 100% open to 0% open

Note: F. STOP and A. STOP levers will not work if

in **DF** mode

- 11. Focus on specimen by adjusting the *Coarse/Fine Focus* knobs
- 12. Switch to higher magnification objectives if desired by rotating nosepiece
- 13. Repeat steps 7-12 until desired magnification and image quality is obtained
- 14. Go to **Step VIII. Image Capture** when ready to acquire image



VI. EPI: Differential Interference Contrast – 1/2

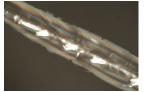
1. Press the **EPI/DIA** selector and set to **EPI**



- 2. Adjust Bright/Dark Field selector lever to desired =
- 5. Push the *Analyzer Plate* in **⇒**

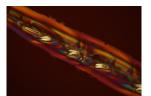


- 7. Rotate the polarizer to adjust the polarization from
 - lateral \rightarrow to vertical \rightarrow
- 8. Push the **DIC Prism** in and set to **Position A**:
- 9. Rotate small knob to adjust contrast and color



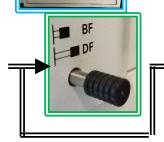




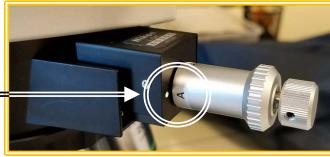












VI. EPI: Differential Interference Contrast – 2/2

10. Select NCB filter (balances color) if desired
11. Adjust the brightness with the Brightness Control → (aperture diaphragm) by sliding levers up and down from 100% open to 0% open

Lower

Stage

Course Focus

Fine Focus

Note: F. STOP and A. STOP levers will not work if in DF mode

- 13. Focus on specimen by adjusting the *Coarse/Fine Focus* knobs
 14. Switch to higher magnification objectives if desired by rotating
- 15. Repeat steps 7-14 until desired magnification and image quality is obtained

nosepiece

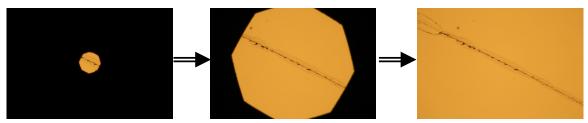
16. Go to Step VIII. Image Capture when ready to acquire image

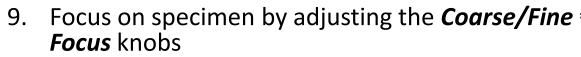
VII. DIA: Bright Field – 1/2

Press the **EPI/DIA** selector and set to **DIA** Push *Bright/Dark Field* selector lever to fully in *BF* position Select NCB filter (balances color) if desired Adjust the brightness with the *Brightness Control* \Rightarrow Adjust the Field Diaphragm Control to fully closed Adjust the **Condenser Height** = until the field diaphragm is focused

VII. DIA: Bright Field – 2/2

- 7. Center the field diaphragm by adjusting *Centering Screws*
- 8. Open the *Field Diaphragm Control* until field diaphragm circumscribes the field of view







$$10X = 0.3$$

$$20X = 0.45$$

$$50X = 0.8$$

$$100X = 0.9$$

- 10. Switch to higher magnification objectives if desired by rotating nosepiece
- 7. Repeat steps 3-11 until desired magnification and image quality is obtained
- 8. Go to Step VIII. Image Capture when ready to acquire image

VIII. Image Capture – 1/1

- 1. Click on the **Folder** icon and select desired folder to store saved pictures in
- Recommend creating you own personal folder with sub-folders for each sample to help distinguish among them later
- 3. It is important to record the objective used for **EACH** image taken (necessary for scale)
- 4. Review *Camera Settings* before acquiring image
- 5. Click on the *Shutter Button* to acquire your image



IX. Cleanup -1/1

1. Lower the stage away from the objectives by rotating the *Coarse Focus* knob **TOWARD** you



3. Turn off the Lamp Power Switch at the back of the microscope

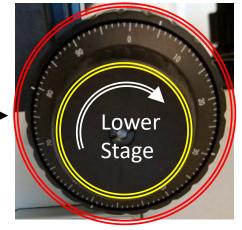
MANUAL

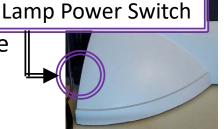


5. Sign-off from your account

6. Clean up and dispose of any consumables used and return any tools back to its respective containers or bins

7. Confirm that the microscope is turned **OFF** again (**NO LIGHT!**), then place cover over microscope







X. ImageJ - 1/3

1. Double-click on *ImageJ* icon

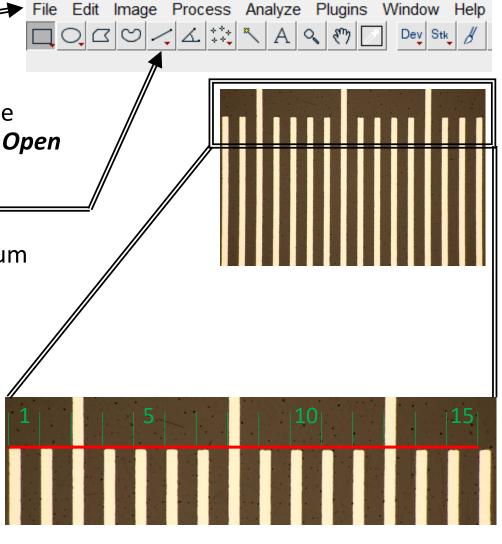


↓ ImageJ

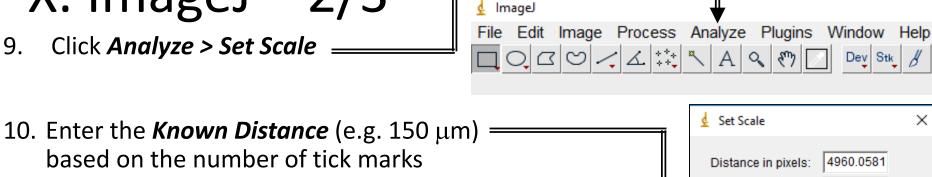
- 2. Click *File > Open*
- 3. Locate the *Scale Bar Images* folder
- Select the *Magnification* of the image you wish to measure (e.g. 100X) and *Open*
- 5. Click the **Segment Tool** and select **Straight Line**
- Draw a line that contains the maximum number of tick marks

Note: It matters where you start and end the line!

- 7. Count the number of tick marks contained (e.g. 15)
- 8. Each division is 0.01 mm (or 10 μ m)



X. ImageJ - 2/3



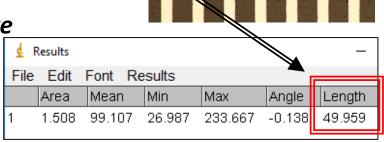
11. Enter the *Unit of Length* to desired unit (e.g. mm)

and each division = 0.01 mm (or 10 μ m)

- 12. Check *Global* to set scale for all images =
- 13. Confirm your scale by drawing a new *Straight Line* —
- 14. Click **Analyze > Measure** and check value

If incorrect, repeat steps 5 - 13

- 15. Click *File > Open* and select your image(s) of interest
- 16. Draw *Straight Lines* and click *Analyze > Measure*
- 17. Repeat steps 4 16 for other *Magnifications*



Known distance:

Pixel aspect ratio:

Unit of length:

✓ Global

Scale: 33.0671 pixels/µm

150

1.0

|µm

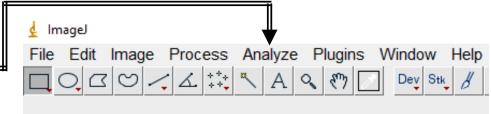
Click to Remove Scale

Cancel

Help

X. ImageJ - 3/3

9. Click *Analyze > Tools > Scale Bar*



- 10. Enter <u>Width in um</u> (e.g. 50 μ m) based on the length of scale bar desired
- 11. Enter *Height in pixels* for desired scale bar thickness
- 12. Enter *Font size* for desired text size
- 13. Identify *Color* of the scale bar
- 14. Identify *Background* color (if desired)
- 15. Identify *Location* where *Scale Bar* to be placed

